

S-cell Problem (home work)

- Calculate velocity field in an S-cell (3D fluidic cell, 50 μm height).
- What would flow field uniformity across a 1mmx1mm array spotted in the middle of the cell?

Data:

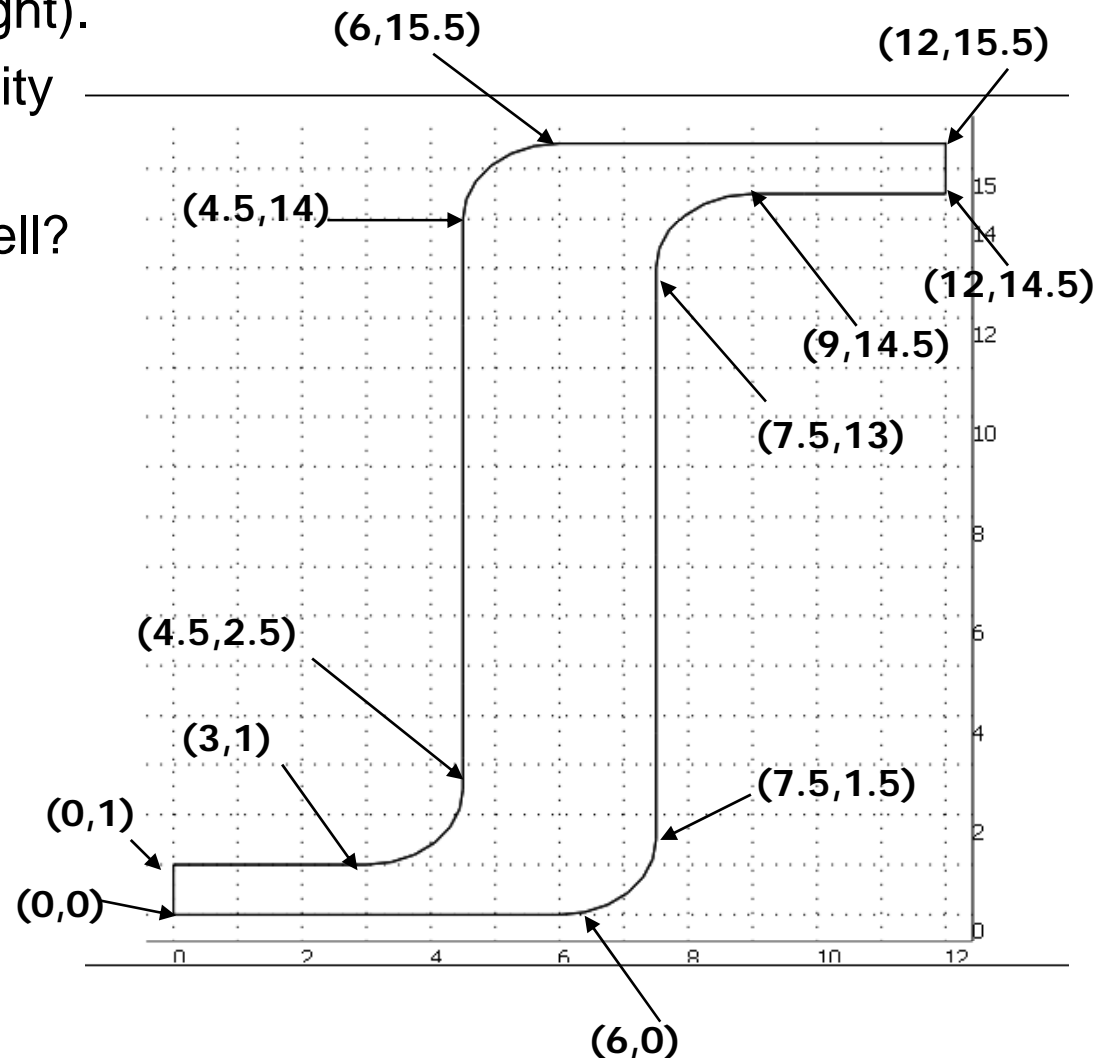
Flow rate: 100 $\mu\text{l}/\text{min}$

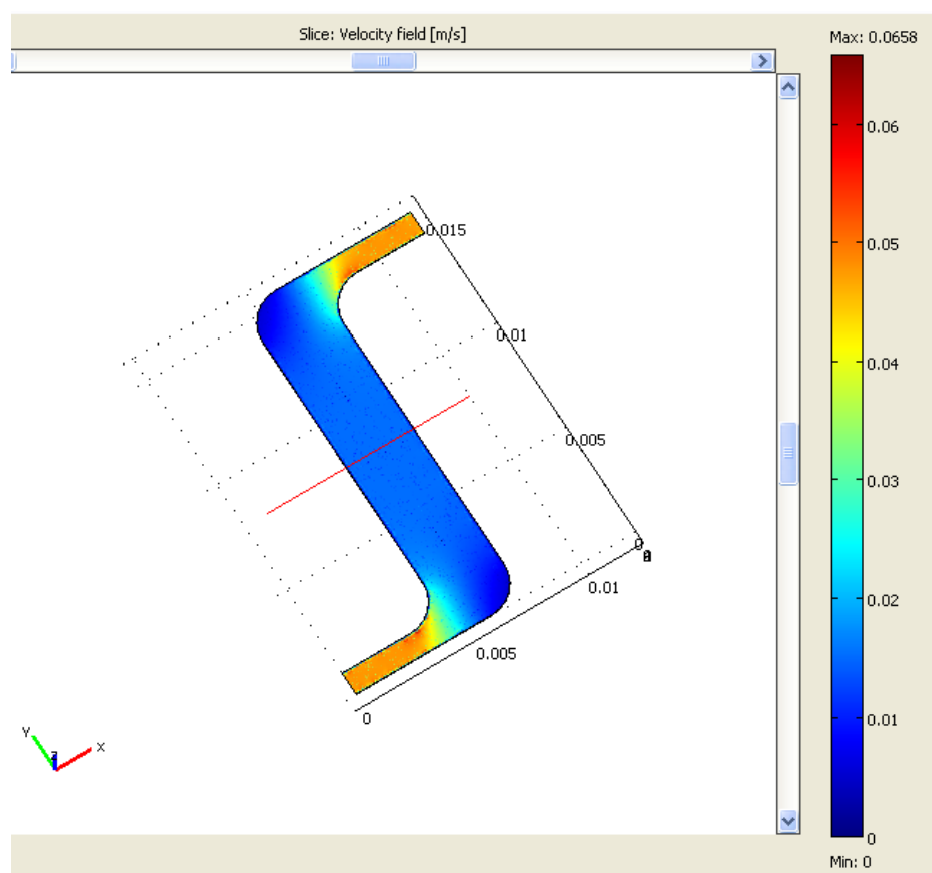
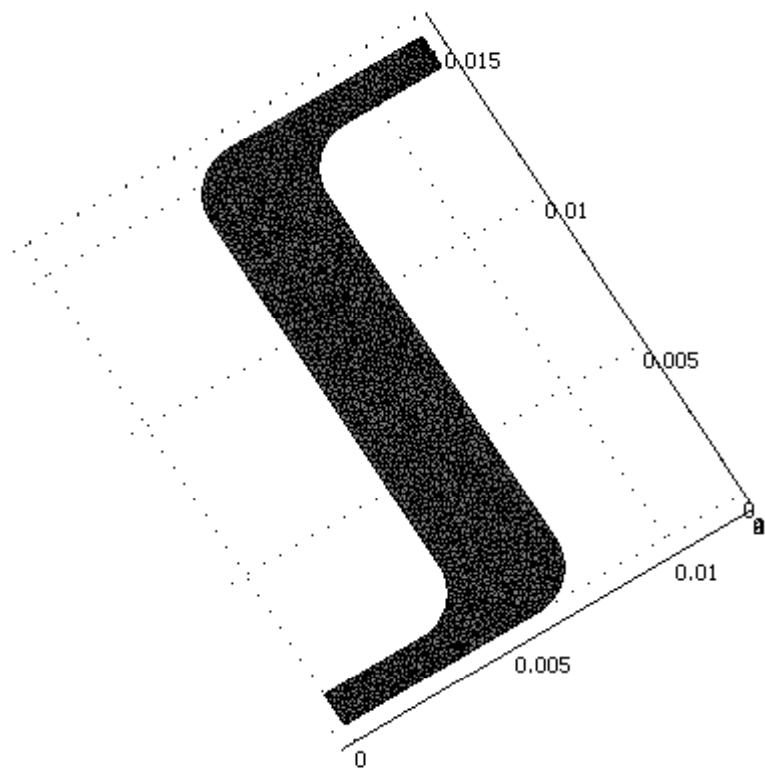
Liquid: water, $T=298\text{K}$

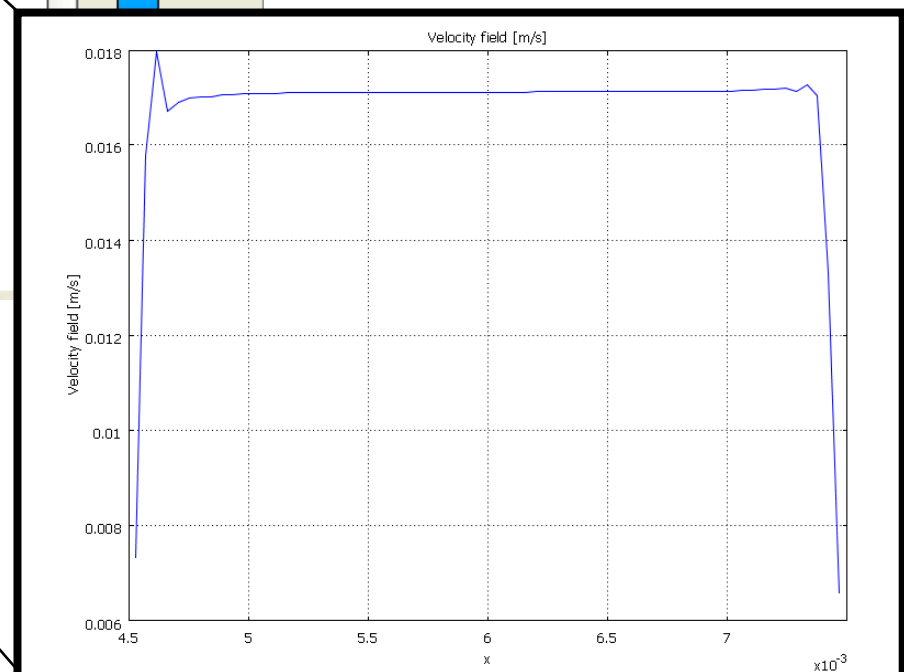
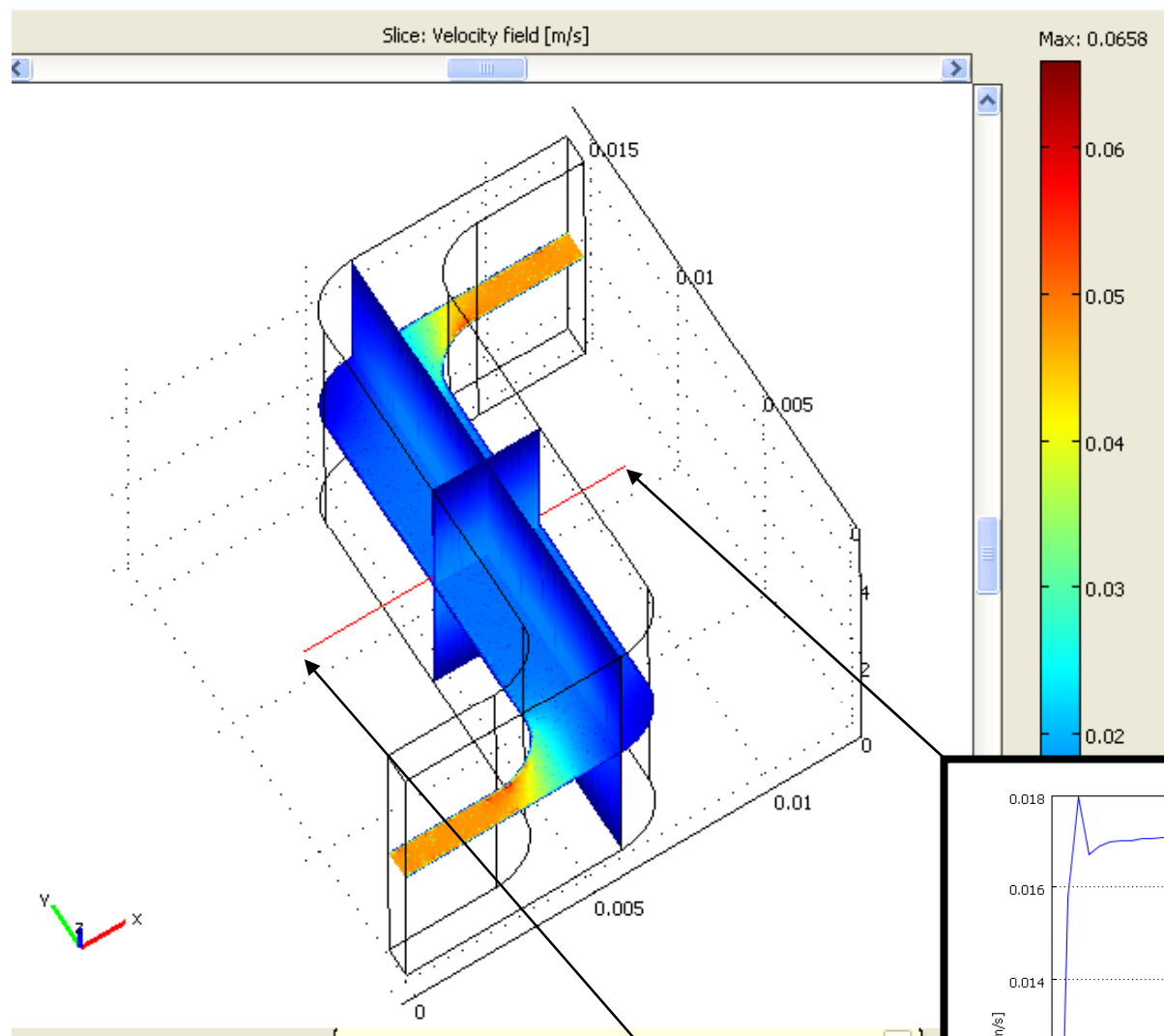
Channel height (z): 50 μm

Channel width: 1mm in/outlet;
3mm chamber

Curvature radius: 1.5mm





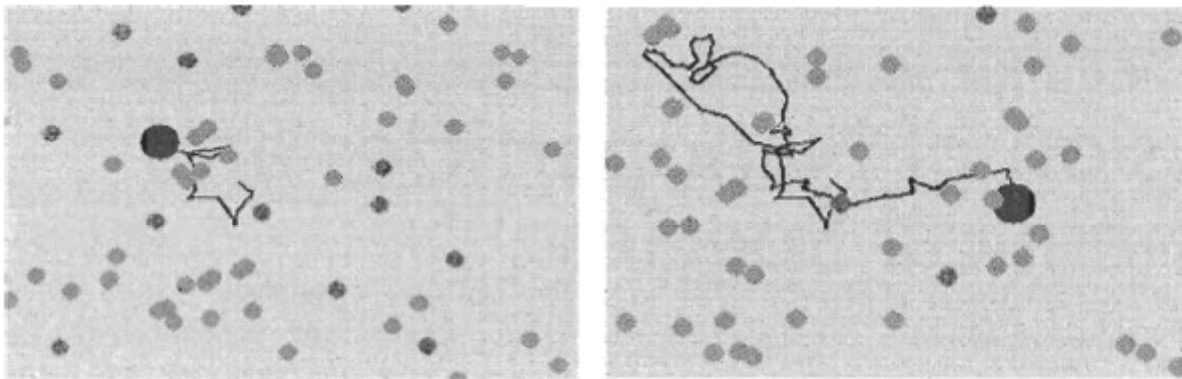


Lecture 7

**Flow and Diffusion.
Micromixers. Multiphysics
modelling with COMSOL**

Brownian motion

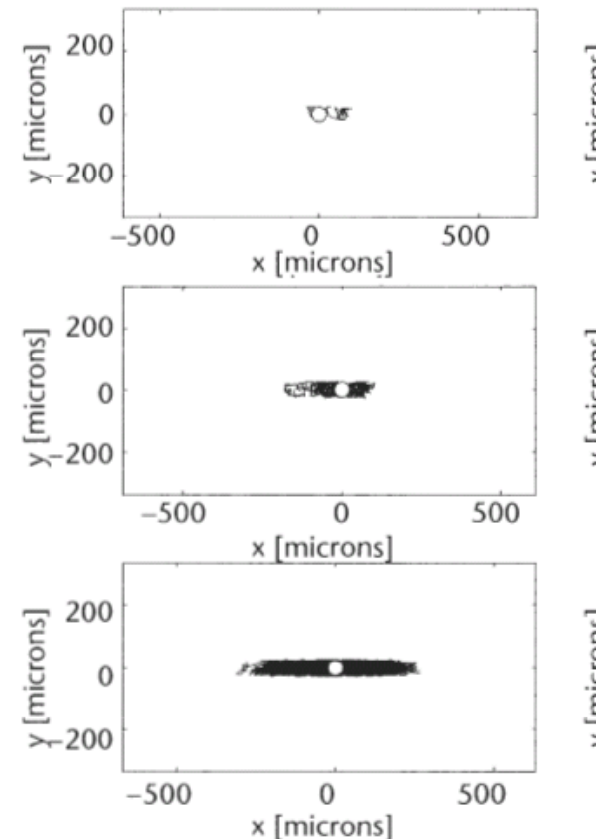
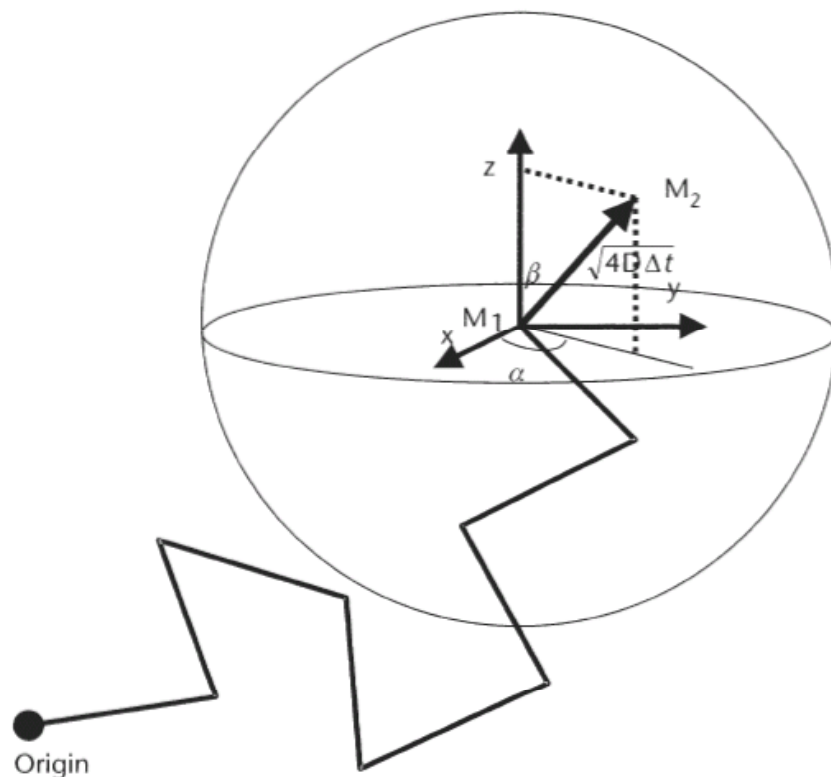
- discovered by R. Brown and J.Ingenhous by observation of pollen grains floating on water



- macroscopic (concentration) and microscopic approach to diffusion

Random walk

- Diffusion can be modeled as a random walk using Monte-Carlo simulation



Microscopic approach

- Model: 1D random walk on a grid (uncorrelated steps left and right with constant step length l)

$$\langle \Delta x_i \cdot \Delta x_j \rangle = l \delta_{ij}$$

- Average distance after N steps (averaged over M walks):

$$\langle \Delta x_N \rangle = \frac{1}{M} \sum_{j=1}^M x_N^{(j)} = \frac{1}{M} \sum_{j=1}^M \sum_{i=1}^N x_i^{(j)} = \sum_{i=1}^N \langle x_i \rangle = 0$$

- Rms distance after N steps (averaged over M walks):

$$\begin{aligned} \langle \Delta x_N^2 \rangle &= \frac{1}{M} \sum_{j=1}^M \left[x_N^{(j)} \right]^2 = \frac{1}{M} \sum_{j=1}^M \left(\sum_{i=1}^N x_i^{(j)} \right) \left(\sum_{k=1}^N x_k^{(j)} \right) = \\ &= \frac{1}{M} \sum_{j=1}^M \left(\left(\sum_{i=1}^N \left[x_i^{(j)} \right]^2 \right) + \underbrace{\left(\sum_{i=1}^N \sum_{k \neq i}^N x_i^{(j)} x_k^{(j)} \right)}_{=0} \right) = Nl^2 + \sum_{i=1}^N \sum_{k \neq i}^N \langle x_i^{(j)} x_k^{(j)} \rangle \end{aligned}$$

Microscopic approach (cont)

- Rms distance after N steps

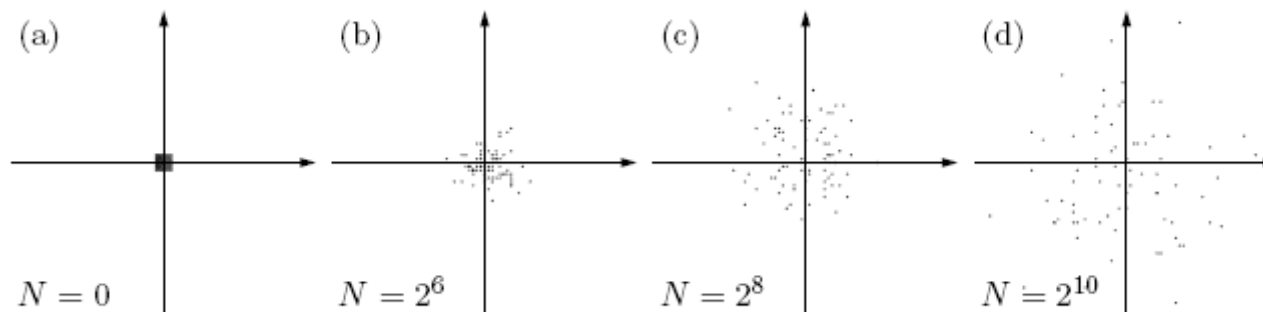
$$\langle \Delta x_N^2 \rangle = Nl^2$$

$$l_{diff,N} = \sqrt{\langle \Delta x_N^2 \rangle} = \sqrt{N}l = \sqrt{\frac{tl^2}{\tau}} = \sqrt{Dt}$$

- In case of uncorrelated 2D steps

$$\langle R_N^2 \rangle = \langle x_N^2 + y_N^2 \rangle = \langle x_N^2 \rangle + \langle y_N^2 \rangle = 2Nl^2$$

$$l_{diff}^{2D} = \sqrt{2Dt}$$



Macroscopic approach to diffusion

- First Fick's law

$$J = -D\nabla c$$

- Second Fick's law

$$\frac{\partial c}{\partial t} = D\Delta c$$

$$\frac{\partial c}{\partial t} = D\Delta c - v\nabla c + S$$

source/sink term

- Einstein formula

$$D = \frac{kT}{C_D}$$

$$D = \frac{kT}{6\pi\eta R}$$

Convection-diffusion equation

- Continuity equation for heterogeneous fluid

$$\frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \vec{v}) = 0; \quad \rho(\vec{r}) \equiv \sum_a \rho_a(\vec{r}); \quad C_\alpha(r, t) \equiv \frac{\rho_\alpha(r, t)}{\rho(r, t)}$$

- in integral form

$$\int_{\Omega} \frac{\partial C_\alpha \rho}{\partial t} dV = - \int_{\partial \Omega} n \cdot (C_\alpha \rho \vec{v}(r, t) + \vec{J}_\alpha^{diff}) da \stackrel{\text{Gauss theorem}}{=} - \int_{\Omega} \nabla \cdot (C_\alpha \rho \vec{v}(r, t) + \vec{J}_\alpha^{diff}) dV$$

$$\frac{\partial C_\alpha \rho}{\partial t} = \nabla \cdot (C_\alpha \rho \vec{v}(r, t) + \vec{J}_\alpha^{diff})$$

$$\rho \left[\frac{\partial C_\alpha \rho}{\partial t} + \vec{v} \cdot \nabla C_\alpha \right] = - \nabla \cdot \vec{J}_\alpha^{diff}$$

$$\frac{\partial C_\alpha}{\partial t} + \vec{v} \cdot \nabla C_\alpha = D_\alpha \nabla^2 C_\alpha$$

convection-diffusion equation

Diffusion equation

- diffusion equation $\frac{\partial c}{\partial t} = D \nabla^2 c$
- from dimensional analysis $L_0 = \sqrt{DT_0}$
- Typical values:
 - $D \approx 2 \cdot 10^{-9} \text{ m}^2/\text{s}$, small ions in water
 - $D \approx 5 \cdot 10^{-10} \text{ m}^2/\text{s}$, sugar molecules in water
 - $D \approx 4 \cdot 10^{-11} \text{ m}^2/\text{s}$, 30bp DNA in water
 - $D \approx 1 \cdot 10^{-12} \text{ m}^2/\text{s}$, 5kbp DNA in water

Limited point-source diffusion

- Normal (Gaussian) distribution

$$P(s) \equiv \frac{1}{\sqrt{2\pi}} e^{-\frac{1}{2}s^2}; \langle s \rangle = \int_{-\infty}^{\infty} sP(s)ds = 0; \langle s^2 \rangle = \int_{-\infty}^{\infty} s^2 P(s)ds = 1;$$

- Let's consider diffusion from a point source described by δ -function. $c(x, 0) = N_0 \delta(x)$

The solution of the diffusion equation is:

$$c(x, t) = N_0 (4\pi Dt)^{-1/2} \exp\left[-\frac{x^2}{4Dt}\right] = N_0 P(s_x), \quad s_x^2 = \frac{x^2}{2Dt}$$

$$l_{diff}^2 = \langle x^2 \rangle = 2Dt \langle s_x^2 \rangle = 2Dt$$

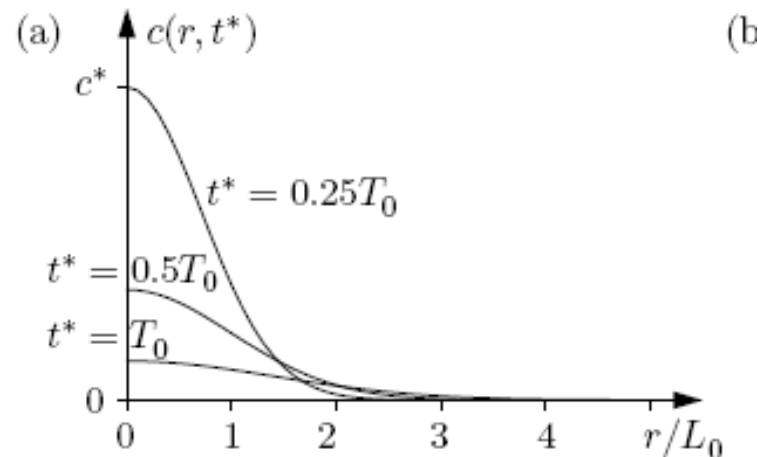
Limited point-source diffusion

- In 2D case:

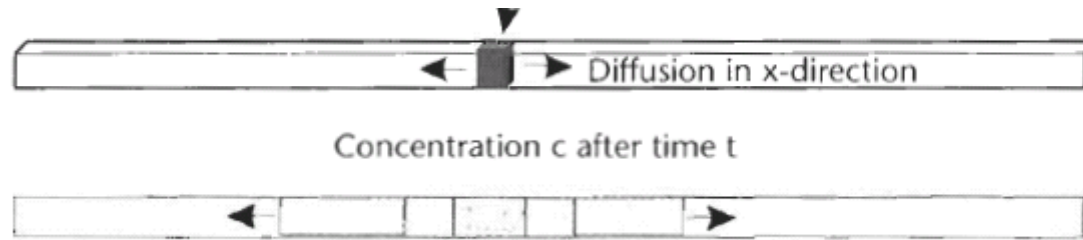
$$c(x, y, 0) = N_0 \delta(x) \delta(y)$$

$$c(x, t) = N_0 (4\pi Dt)^{-1/2} \exp\left[-\frac{x^2}{4Dt}\right] \times (4\pi Dt)^{-1/2} \exp\left[-\frac{y^2}{4Dt}\right] =$$
$$= N_0 P(s_x) P(s_y)$$

$$l_{diff, 2D}^2 = \langle x^2 + y^2 \rangle = 2Dt \langle s_x^2 + s_y^2 \rangle = 4Dt$$

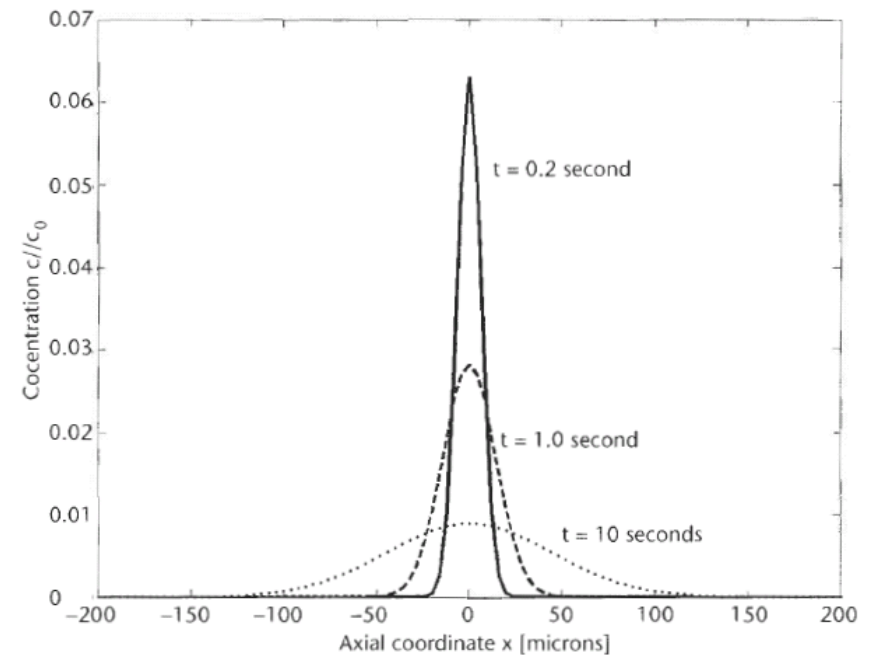


Spreading from a point source in 1D



- solution:

$$c(x, t) = \frac{c_0}{\sqrt{4\pi Dt}} e^{-\frac{x^2}{4Dt}}$$



Limited planar source

- Let's consider diffusion into half space.

$$c(r, 0) = 2N_0\delta(x)$$

$$c(r, t) = N_0(\pi Dt)^{-1/2} \exp\left[-\frac{x^2}{4Dt}\right] = 2N_0P(s_x), \quad s_x^2 = \frac{x^2}{2Dt}$$

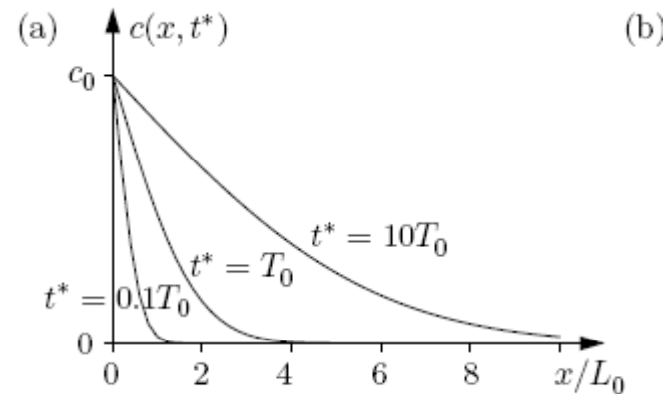
Constant planar source diffusion (Ilkovic's solution)

- Problem: consider a half-space with an initial concentration c_0 . Concentration on the wall is zero at any time. Find the concentration profile vs time.

$$c = c_0 \operatorname{erf} \left(\frac{x}{\sqrt{4Dt}} \right) \quad \operatorname{erf}(x) = \frac{2}{\sqrt{\pi}} \int_0^{\infty} e^{-u^2} du$$

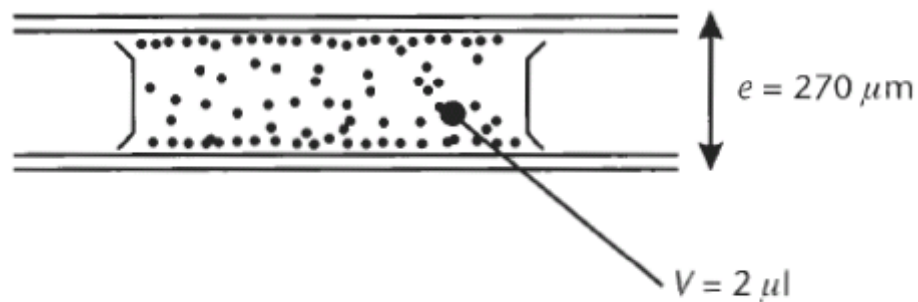
$$\frac{\partial c}{\partial x} = c_0 \frac{2}{\sqrt{\pi}} e^{-\frac{x^2}{4Dt}} \frac{1}{\sqrt{4Dt}}$$

$$J = -D \nabla c|_{\text{wall}} = -c_0 \sqrt{\frac{D}{\pi t}}$$



Example: diffusion between two plates

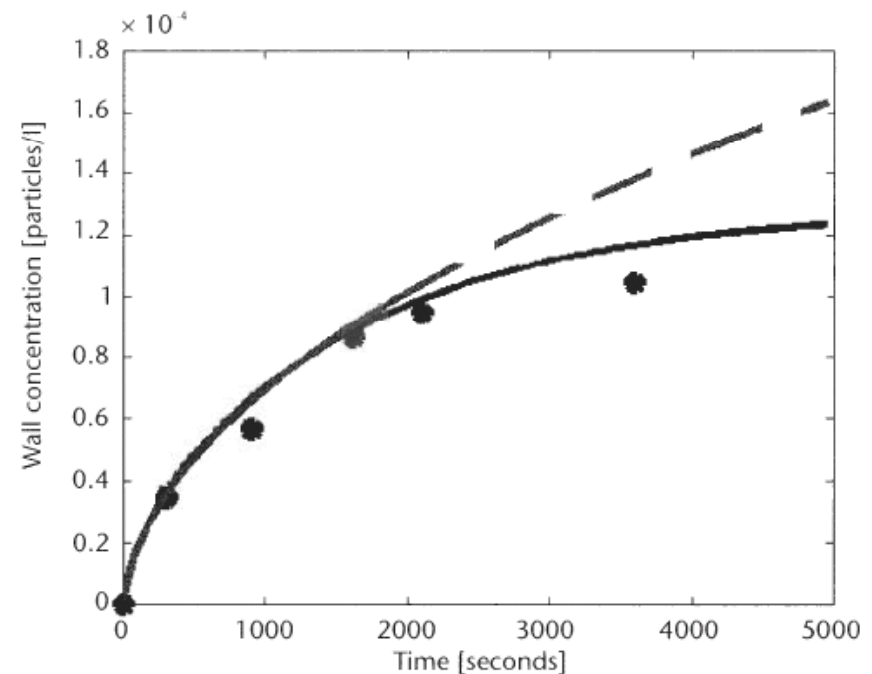
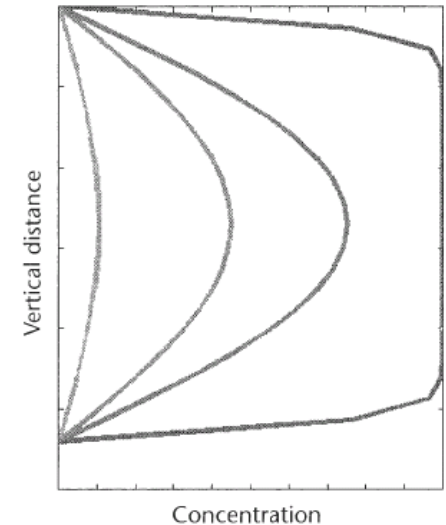
- A liquid drop contains nanoparticles that are immobilized upon contact with the walls. Find the concentration dependence vs time



- For times less than $\tau = \frac{e/2}{4D}$

Ilkovic's solution can be used:

$$J = -2c_0 \sqrt{\frac{D}{\pi t}}$$



Navier-Stokes equation: Diffusion of momentum

$$\rho \frac{\partial \vec{v}}{\partial t} = -\nabla p + \rho \vec{g} + \mu \nabla^2 \vec{v}$$

- In the case of decelerating Poiseuille flow

$$\frac{\partial \vec{v}}{\partial t} = \left(\frac{\mu}{\rho} \right) \nabla^2 \vec{v}$$

kinematic viscosity ν

Diffusion constant for momentum!

- Characteristic time for diffusion across the channel

$$T = \frac{a^2 \rho}{\mu}$$

$$\text{Schmidt number} = Sc = \frac{\nu}{D} = \frac{\mu}{\rho D}$$

intrinsic property of the solution

Diffusion vs Sedimentation

Does the gravity force affects the diffusion?

- Let's compare diffusion and sedimentation time across a microfluidic chamber

- sedimentation time: $C_D V_s = 6\pi\eta R_H V_s = \Delta\rho g V_p$

$$V_s = \frac{2}{9} \frac{\Delta\rho g R^2}{\eta}$$

$$\tau_1 = d / V_s$$

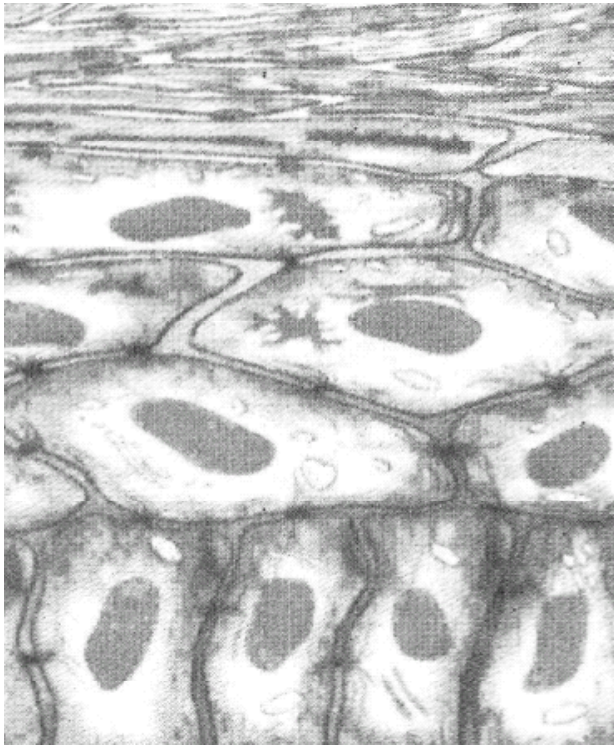
- diffusion time: $\tau_2 = \frac{d^2}{4D}$

$$\beta = \frac{\tau_1}{\tau_2} = \frac{d}{V_s} \frac{4D}{d^2} = 4 \frac{kT}{\Delta m g} \frac{1}{d}$$

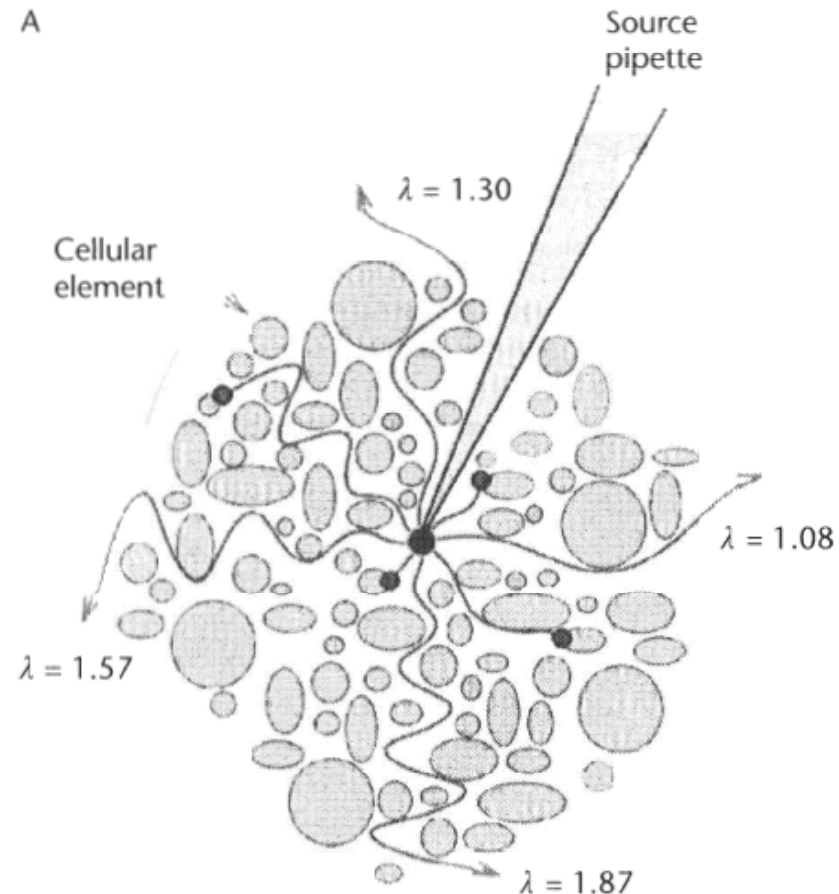
- if $\beta \ll 1$ sedimentation dominates

Diffusion in confined volumes

- For example, delivery of drugs relies on a diffusion in ECS of cellular clusters ^A



cell arrangement in the human skin

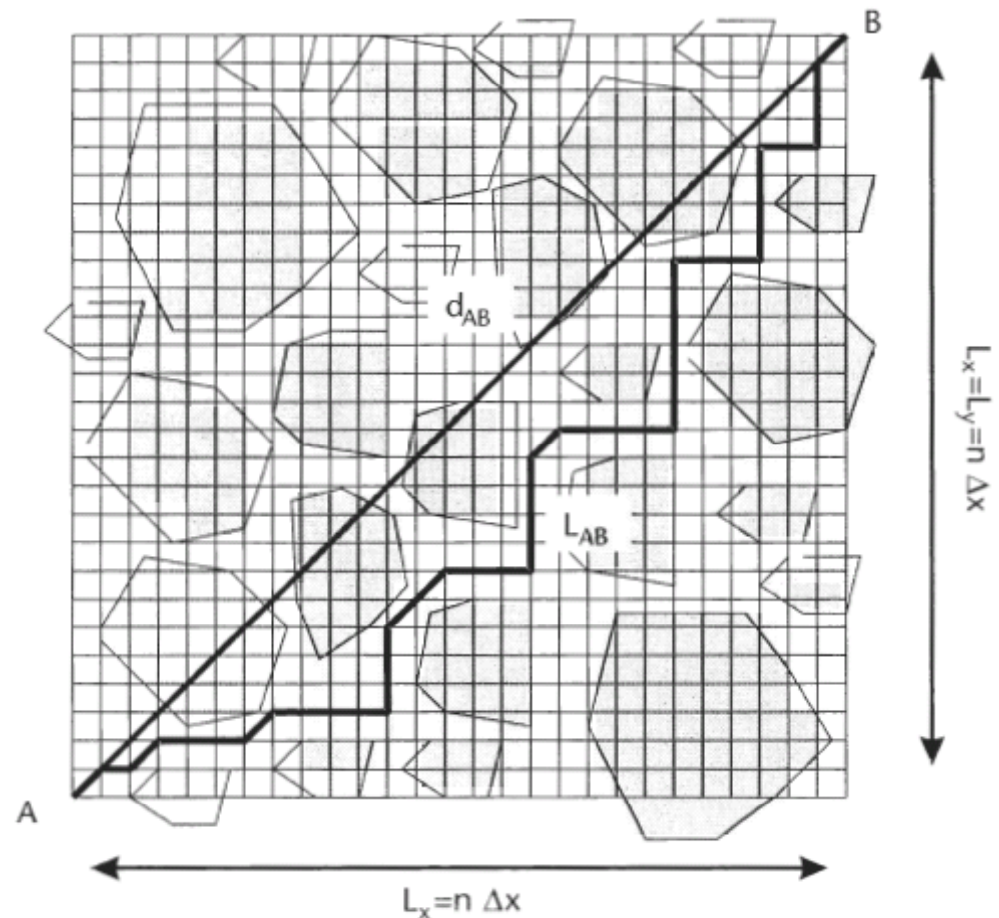
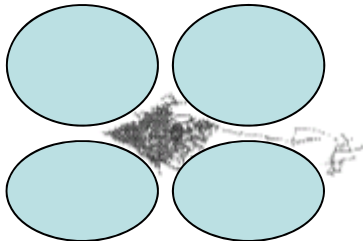


- tortuosity: ration between the distance in liquid and the straight distance between the points

Diffusion in confined volumes

It can be shown that:

- for any 2D regular isotropic lattice tortuosity is equal to $\tau = \sqrt{2}$
- for 3D: $\tau = \sqrt{3}$
- The situation is more complicated for irregular cells and in the presence of intercleft volumes

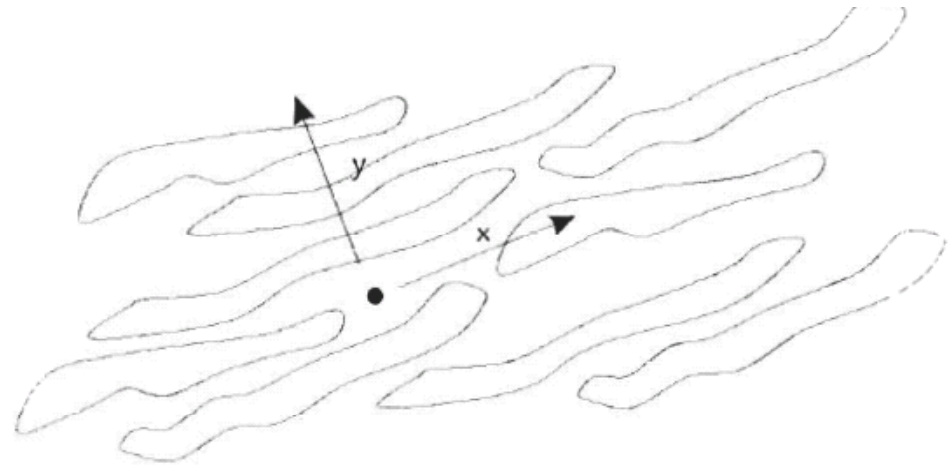


How to treat anisotropic media

- to treat a media with a preferential direction we have to introduce a diffusion tensor:

$$[D] = \begin{bmatrix} D_{11} & D_{12} & D_{13} \\ D_{21} & D_{22} & D_{23} \\ D_{31} & D_{32} & D_{33} \end{bmatrix}$$

$$\begin{bmatrix} J_1 \\ J_2 \\ J_3 \end{bmatrix} = -[D] \nabla c$$

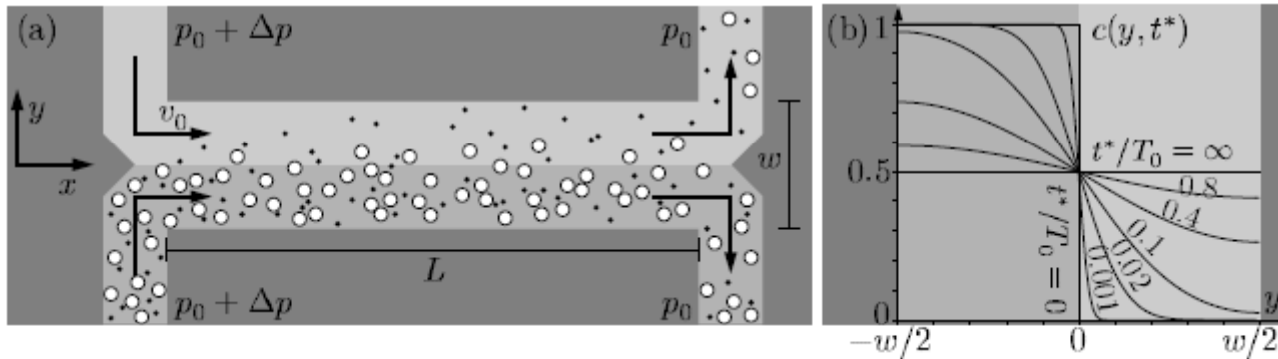


$$\begin{aligned} \frac{\partial c}{\partial t} = & D_{11} \frac{\partial^2 c}{\partial x^2} + D_{22} \frac{\partial^2 c}{\partial y^2} + D_{33} \frac{\partial^2 c}{\partial z^2} + (D_{23} + D_{32}) \frac{\partial c}{\partial y \partial z} \\ & + (D_{31} + D_{13}) \frac{\partial c}{\partial z \partial x} + (D_{12} + D_{21}) \frac{\partial c}{\partial x \partial y} \end{aligned}$$

- by rotating and scaling the coordinates it's possible to return to a isotropic (scalar) D:

$$\frac{\partial c}{\partial t} = D \left[\frac{\partial^2 c}{\partial \xi^2} + \frac{\partial^2 c}{\partial \eta^2} + \frac{\partial^2 c}{\partial \zeta^2} \right]$$

H-filter: separating solutes by diffusion



- H-filter takes advantages of laminar flow (flow don't mix) and fast diffusion over short distances

- Characteristic times:

convection time: $\tau_{conv} = \frac{L}{v_0}$

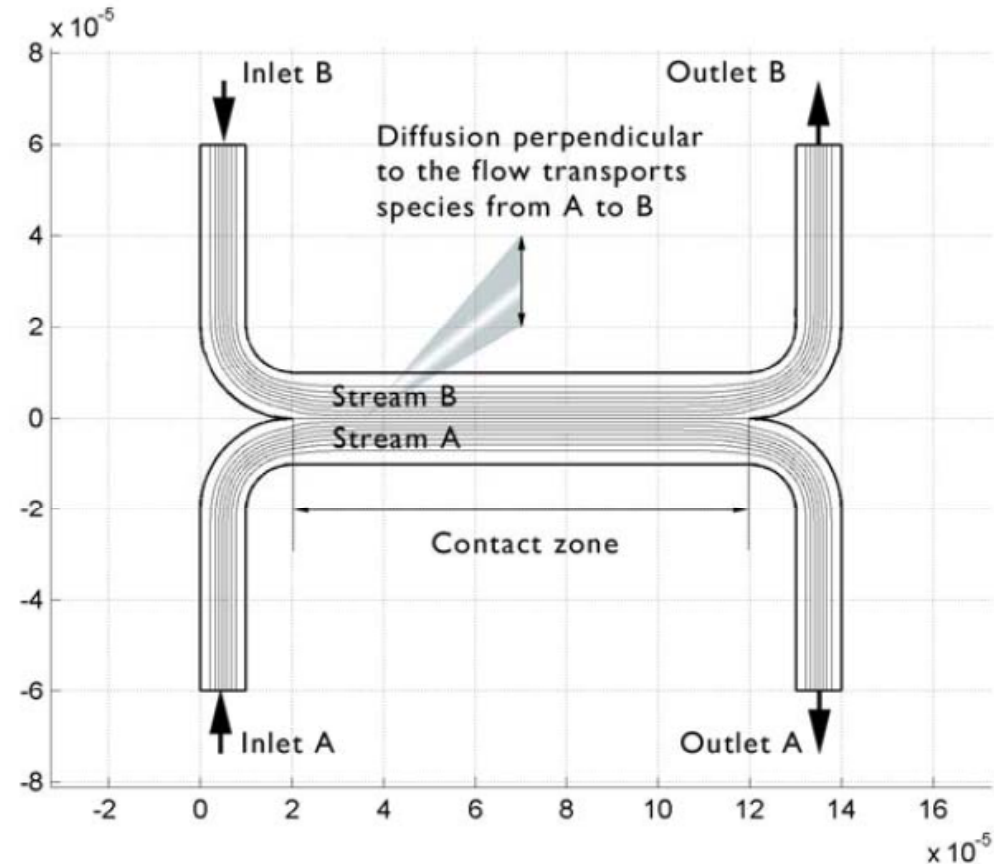
diffusion time: $\tau_{diff} = \frac{w^2}{4D}$

$\tau_{conv} \geq \tau_{diff}$ concentration of solute is the same through the channel

- Critical diffusion constant: $D^* = \frac{v_0 w^2}{4L}$

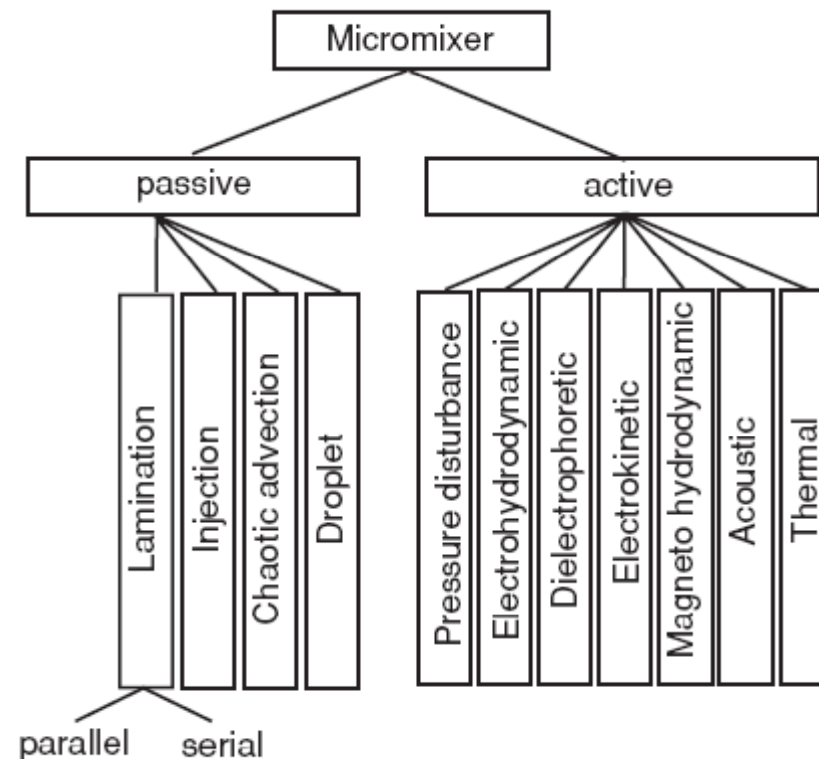
H-cell model

- H-cell perform separation via diffusion during controlled time:
 - Two laminar streams are brought into contact for given amount of time
- Small species from A can diffuse into B
- Modelling parameters:
 - Cell height – $20\mu\text{m}$
 - Pressure at inlets: $P_0=2\text{ Pa}$;
- Calculate critical diffusion constant and compare with the modelling results



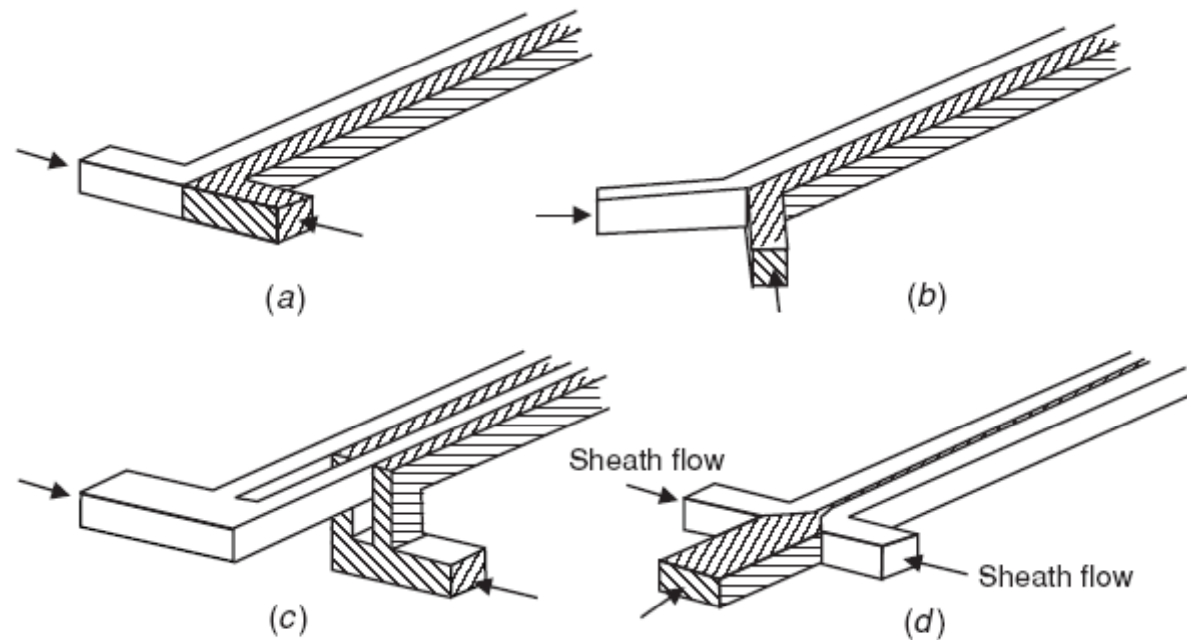
Mixing on microscale

- Flow in microchannels is laminar and mixing occurs through diffusion (slow!)
 - Chemical reactions (necessary for Lab-on-Chip assays, but also for DNA and protein synthesis etc.) require mixing of reagents
-



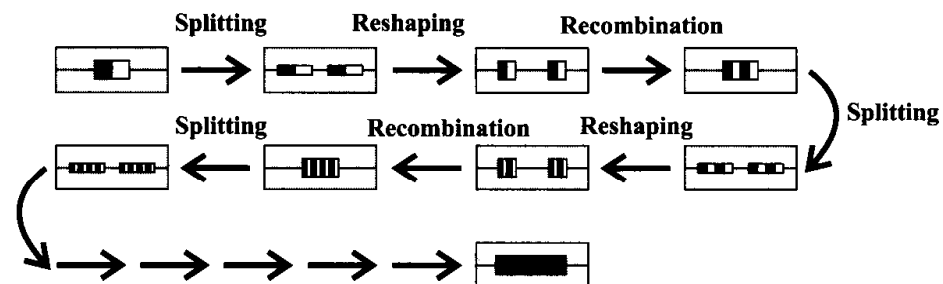
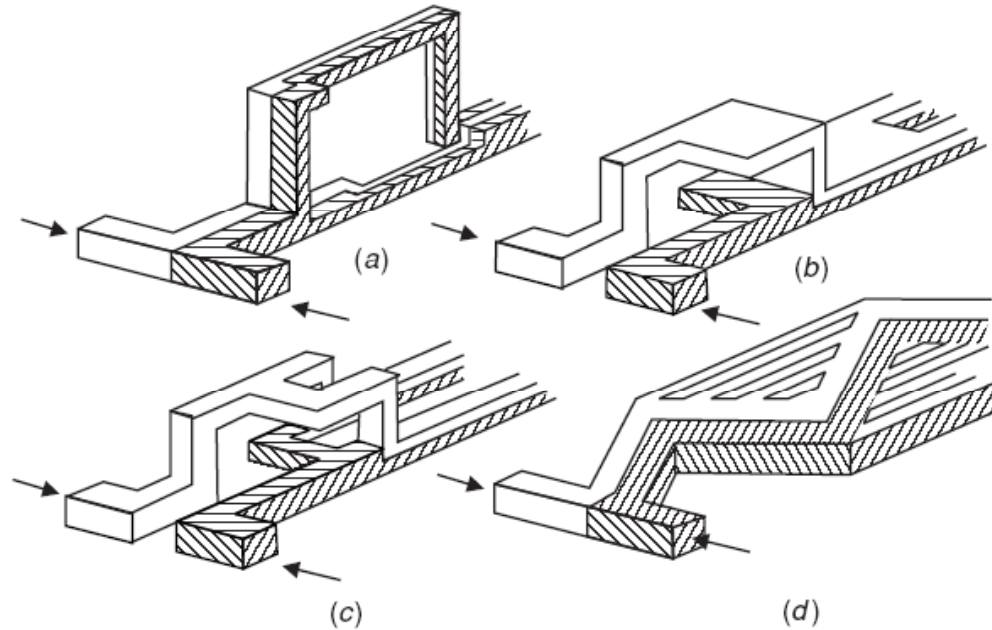
Passive micromixers

- General approach: to shorten diffusion time via increased contact/decreased thickness or by chaotic advection
-
- Parallel lamination:



Passive micromixers

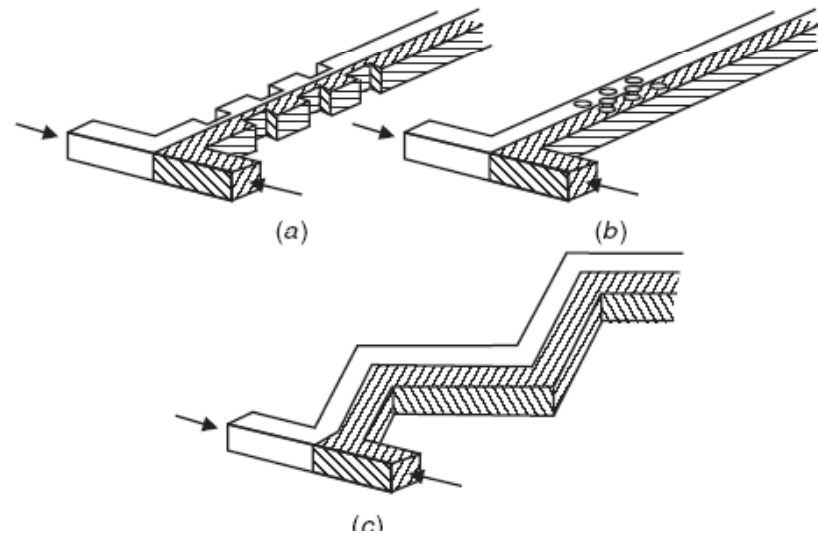
- Serial lamination:



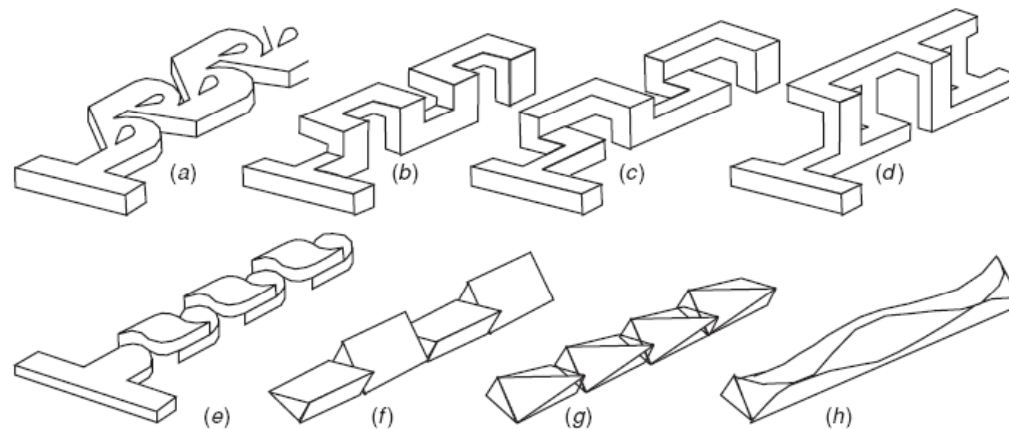
Passive micromixers

- Chaotic advection at high Re numbers

Recirculation are produced at the zig-zag turns of channel or behind the obstacles. Critical Re numbers are reported ~ 80 for zig-zag channel

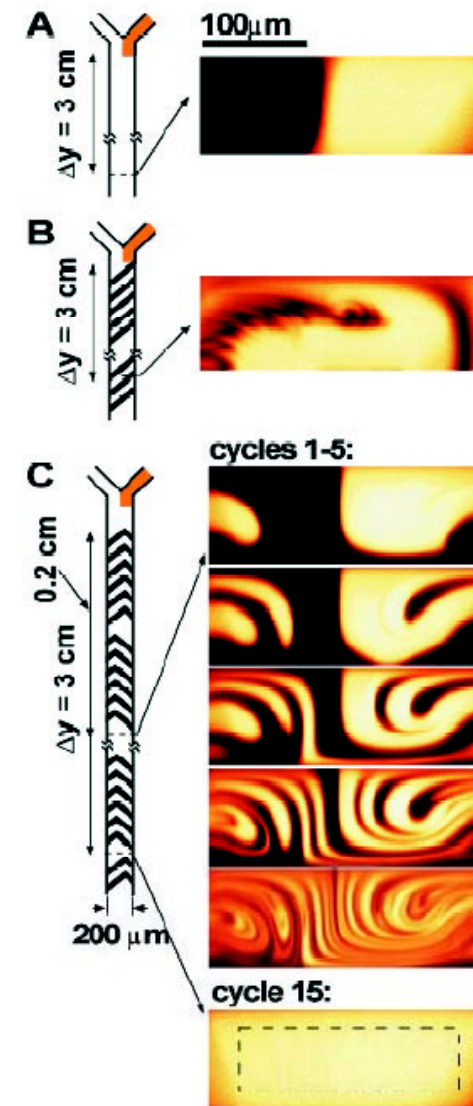


- Chaotic advection at intermediate Re numbers



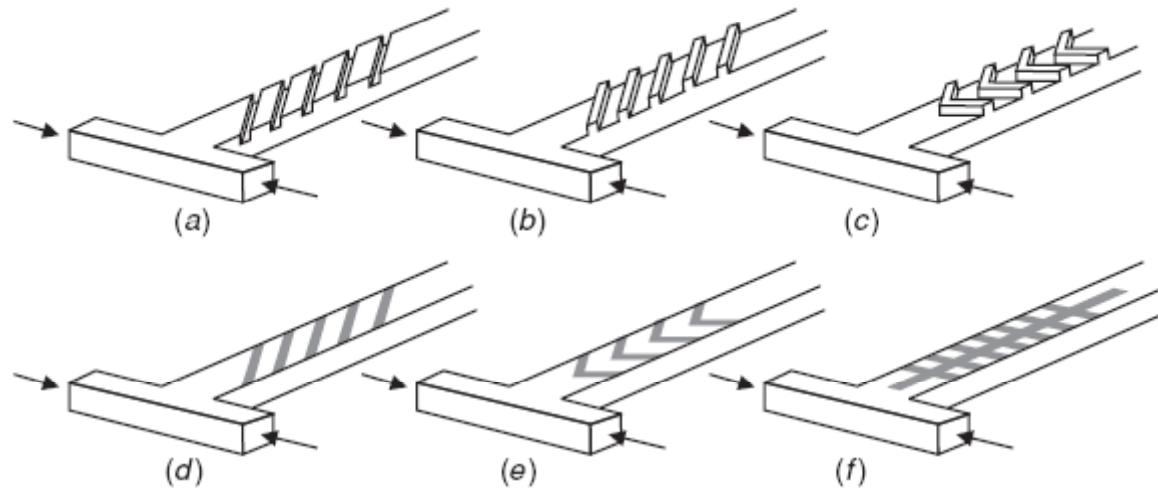
Passive micromixers

- Another way to induce mixing in laminar flow – patterned surface
 - Top -- no surface patterning
 - Middle – slants
 - Bottom V-shape (herringbone) – clearly see enhanced mixing
- Effective at low Reynolds numbers (in the range of 1- 100)



Passive micromixers

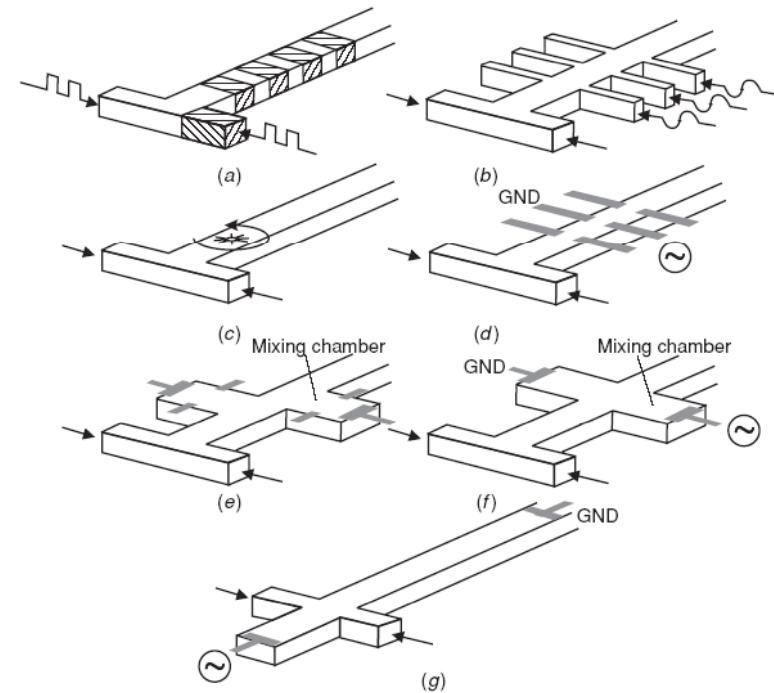
- Design variations of chaotic advection mixers



- Droplet micromixers

Active micromixers

- Function via
 - pressure disturbance,
 - mechanical microstirrers,
 - electrohydrodynamic
 - electrophoretic disturbance



Problems

- An analyte ($c_a=1$) is injected into S-cell. Plot concentration vs. time in the middle of the cell. How concentration is varied across the 1mm central area?
- Problems 5.7 and 5.8 (Bruus).

